

following manner. A 5.3-g. sample, m.p. 156–163°, was treated with a mixture of 15 cc. of carbon disulfide and 5 cc. of nitromethane, whereupon all but 1.92 g. of solid dissolved. The insoluble portion was crystallized twice from acetone to yield 0.70 g. of 2,2,5,5-tetraphenyltetrahydrofuran (I). Recrystallizations of the soluble portion of the original mix-

ture eventually yielded 0.12 g. of 1,1,4,4-tetraphenyl-1,3-butadiene (III) as the only other compound which could be isolated in pure condition.

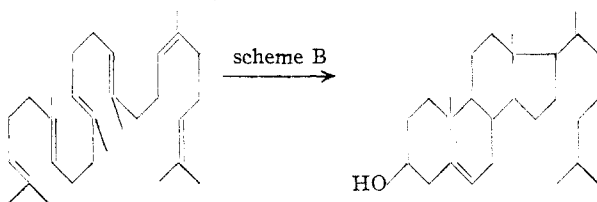
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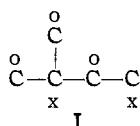
ON THE INCORPORATION OF ACETATE INTO CHOLESTEROL

Sir:

It has recently been demonstrated¹ that squalene is an efficient precursor of cholesterol and is most likely an intermediate in the conversion of acetate to this sterol. A pathway by which this hydrocarbon could be transformed into cholesterol was suggested by Robinson² in 1934 (route A). The recent findings^{3,4} that the sterols and tetracyclic triterpenes are most likely of the same absolute configuration have suggested an alternate mechanism for the utilization of squalene in the biosynthesis of cholesterol (scheme B).



Degradation of the side-chain⁵ and of ring A⁶ of cholesterol formed from C¹⁴-labeled acetate in biological experiments has indicated that such a biosynthesis might proceed by condensation of isoprenoid units labeled as shown in (I).⁷ Therefore, if route A was followed



carbon atoms 10, 13, 20 and 25 of cholesterol would be derived from a carboxyl carbon of acetate and carbon atoms 18, 19, 21 and 26 from the methyl carbon. Such would not be the case if the tetracyclic triterpenoid type of scheme (B) was followed since a migration of a carbon atom would be involved. Thus in route B, carbon atoms 10, 20 and 25 would be from carboxyl carbons and carbon atoms 13, 18, 19, 21 and 26 from methyl carbons.

- (1) R. G. Langdon and K. Bloch, *J. Biol. Chem.*, **200**, 135 (1953).
- (2) R. Robinson, *J. Soc. Chem. Ind.*, **53**, 1062 (1934).
- (3) W. Klyne, *J. Chem. Soc.*, 2916 (1952).
- (4) W. G. Dauben, D. F. Dickel, O. Jeger and V. Prelog, *Helv. Chim. Acta*, **36**, 325 (1953).
- (5) J. Wüersch, R. L. Huang and K. Bloch, *J. Biol. Chem.*, **195**, 439 (1952).
- (6) J. W. Cornforth, G. D. Hunter and G. Popják, *Biochem. J.*, **53**, xxiv (1953).
- (7) The methyl carbon of acetate is denoted by o and the carboxyl carbon by x.

Hence, the acetic acid derived from a Kuhn-Roth oxidation of cholesterol which had been formed biosynthetically from C¹⁴-methyl-labeled acetic acid should contain C¹⁴ in the carboxyl group if route B was followed and not if route A was utilized.

Such an experiment was performed and the acetic acid so obtained was degraded by a Schmidt reaction. It was found that the acetic acid had a specific activity of 55,⁸ the carboxyl carbon had 35 and the methyl carbon had 74. It is seen that the carboxyl carbon atom of acetic acid derived from cholesterol possessed C¹⁴ as would be predicted by route B and thus would rule against the Robinson postulate (route A).

If one assumes that the ratio of methyl carbons to carboxyl carbons in cholesterol derived from acetate⁹ is 15/12 and that 5 of the 8 carbon atoms of the 4 molecules of acetic acid (from the angular positions as discussed above) obtained from the Kuhn-Roth oxidation of such a labeled cholesterol are, in turn, derived from the methyl carbons, then the specific activity of the carbon dioxide obtained by decarboxylation of acetic acid should have a value of only 22. The higher value, 35, can be reconciled by consideration of the involvement of the progenitor, methyl-labeled acetate, in the tricarboxylic acid cycle which would lead to some doubly-labeled acetate. This degree of randomization is of the same order of magnitude as recently reported by Cornforth, Hunter and Popják.⁶

These results strongly indicate that if squalene is directly utilized in the synthesis of cholesterol, a reaction scheme of type B is strongly suggested.¹⁰

(8) All specific activities are expressed as dis./min./mg. BaCO₃.

(9) H. N. Little and K. Bloch, *J. Biol. Chem.*, **183**, 33 (1950).

(10) A recent report by Woodward and Bloch (*THIS JOURNAL*, **75**, 2023 (1953)) has suggested the same reaction scheme.

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RECEIVED MAY 1, 1953

POSSIBLE SIGNIFICANCE OF LACTONES AS INTERMEDIATES IN OXIDATION OF CARBONACEOUS MATERIALS

Sir:

Controlled oxidation of aqueous, alkaline suspensions of bituminous coals results in the conversion of about 50% of the carbon to a mixture of

water-soluble, polycarboxylic acids.¹ Recent studies in this Laboratory of the reaction products from decarboxylation of the copper salts of these acids have resulted in the recovery of the lactone of 2'-hydroxy-2-biphenylcarboxylic acid (6-dibenzo-pyrone), along with mono- and bicyclic aromatic hydrocarbons. This lactone was characterized by melting point, ultimate analysis, and comparison of infrared spectrum, of ultraviolet spectrum, and of properties of the methoxy acid derivative with those of an authentic sample.

The isolation of this lactone from the oxidation products of coal is highly suggestive in connection with oxidation mechanisms of carbonaceous materials. Lactone rings are very sensitive to pH. In an alkaline hydroxide solution, the ring is opened and the resultant hydroxyl and carboxyl groups will undergo the usual reactions of such groups. In an acid medium, a stable six-membered oxygen-containing ring is formed and further attack on interior rings would be expected to be difficult. The presence of lactones as intermediates would furnish a possible explanation for the much higher rates of oxidation of coals in alkaline than in acid media. Nitric acid is a very effective reagent in the primary stages of the oxidation of coals and various forms of carbon, but to complete the oxidation to benzenecarboxylic acids it has been found advantageous to follow the primary nitric acid oxidation with a secondary one in alkaline medium.² The formation of stable lactone rings in the acidic stage would account for such behavior. This lactone of the biphenyl hydroxy acid is relatively insoluble in aqueous sodium carbonate and this fact suggests an explanation for the lower oxidation rates of coal in sodium carbonate compared with sodium hydroxide solutions. The highest methoxyl values for "regenerated humic acids" are obtained by the Wallaschko³ method, where the compound is dissolved in an excess of alcoholic potash before reaction with dimethyl sulfate. One would expect very complete opening of lactone rings under such circumstances. The esters of acids from oxidation of coal have been shown to form adducts with stannic chloride in dilute pentane solutions. This lactone forms such an adduct under identical experimental conditions.

It has been reported⁴ that the rate of reaction of ozone on coal is markedly affected by the presence of water and that the action of this oxidizing agent on "regenerated humic acids" is greatly accelerated if the acids have been previously treated with boiling aqueous alkali. These facts point to a hydrolytic step in the reaction mechanism.

It is well established that the reaction of steam or water with carbon is greatly accelerated by the presence of alkalis; the opening of peripheral lactone rings could be responsible for the effect. It is possible that surface oxygen complexes, such as the C_xO_y of Rhead and Wheeler,⁵ consist in part of

(1) N. W. Franke, M. W. Kiebler, C. H. Ruof, T. R. Savich and H. C. Howard, *Ind. Eng. Chem.*, **44**, 2784-2792 (1952).

(2) B. Juettner, *THIS JOURNAL*, **59**, 208-213 (1937); O. Grossinsky, *Glückauf*, **88**, 988-995 (1950).

(3) N. Wallaschko, *Arch. Pharm.*, **242**, 242 (1904).

(4) C. R. Kinney and L. D. Friedman, *THIS JOURNAL*, **74**, 57-61 (1952).

(5) T. F. E. Rhead and R. V. Wheeler, *J. Chem. Soc.*, **101**, 846 (1918); **108**, 461 (1912).

lactone rings in peripheral positions. That such lactones can be produced by gas phase oxidation has been demonstrated recently by Brooks⁶ who obtained this identical lactone by the air oxidation of phenanthrene in a fluidized catalyst bed at 370°.

(6) J. D. Brooks, *Research*, **5**, 196 (1952).

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RECEIVED MAY 16, 1953

REACTIONS OF ALLYL ALCOHOL-1-C¹⁴

Sir:

Although an allylic rearrangement would be expected to occur when allyl alcohol is transformed into an allyl halide under certain experimental conditions, the extent to which it takes place has not been determined. In the present work allyl alcohol-1-C¹⁴ was converted to radioactive allyl chloride and allyl bromide by different methods, the starting material and final products degraded with ozone, and the amount of rearrangement determined from the specific activity of the formaldehyde-C¹⁴.

By modification of the excellent method of Young and Lane¹ carbon-14 labeled allyl bromide was prepared from allyl alcohol-1-C¹⁴, phosphorus tribromide, and pyridine at -80°. Upon degradation with ozone, the per cent. rearrangement to allyl bromide-3-C¹⁴ was found to be 46%.

Radioactive allyl chloride was made from allyl alcohol-1-C¹⁴ and thionyl chloride by the method of Meisenheimer and Link² and the amount of rearrangement to allyl chloride-3-C¹⁴ was 51%.

Finally, the tosylate of allyl alcohol-1-C¹⁴ was treated with sodium bromide in a suitable solvent and only allyl bromide-1-C¹⁴ resulted indicating no rearrangement.

Further study is in progress with allyl alcohol-1-C¹⁴ and allyl bromide-1-C¹⁴ to determine if in those cases where rearrangement occurs a unimolecular process of replacement involving the formation of a resonating cation is the predominant mechanism.

(1) W. G. Young and J. F. Lane, *THIS JOURNAL*, **59**, 2051 (1937).

(2) J. Meisenheimer and J. Link, *Ann.*, **479**, 211 (1930).

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RECEIVED MAY 11, 1953

ENZYMATIC SYNTHESIS OF D-GLUTAMINE AND RELATED HYDROXAMIC ACIDS

Sir:

The mechanism of the enzymatic interaction of ATP,¹ L-glutamate, and ammonia, yielding ADP, L-glutamine, and inorganic phosphate has been of interest since the reaction was first described by Speck² and by Elliott.³ Elliott,⁴ using a highly purified enzyme from peas, was unable to separate glutamine synthesis from glutamotransferase ac-

(1) Abbreviations employed: ATP = adenosine triphosphate, ADP = adenosine diphosphate, tris = tris-(hydroxymethyl)-amino-methane.

(2) J. F. Speck, *J. Biol. Chem.*, **168**, 403 (1947); **179**, 1367, 1405 (1949).

(3) W. H. Elliott, *Nature*, **161**, 128 (1948).

(4) W. H. Elliott, *J. Biol. Chem.*, **801**, 681 (1953).